

## Antioxidant and anti-inflammatory properties of C-phycoerythrin from blue-green algae

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**Abstract.** *Objective:* Phycocyanin is a pigment found in blue-green algae which contains open chain tetrapyrroles with possible scavenging properties. We have studied its antioxidant properties.

*Materials and methods:* Phycocyanin was evaluated as a putative antioxidant in vitro by using: a) luminol-enhanced chemiluminescence (LCL) generated by three different radical species ( $O_2^-$ ,  $OH^\bullet$ ,  $RO^\bullet$ ) and by zymosan activated human polymorphonuclear leukocytes (PMNLs), b) deoxyribose assay and c) inhibition of liver microsomal lipid peroxidation induced by  $Fe^{+2}$ -ascorbic acid. The antioxidant activity was also assayed in vivo in glucose oxidase (GO)-induced inflammation in mouse paw.

*Results:* The results indicated that phycocyanin is able to scavenge  $OH^\bullet$  ( $IC_{50} = 0.91$  mg/mL) and  $RO^\bullet$  ( $IC_{50} = 76$   $\mu$ g/mL) radicals, with activity equivalent to 0.125 mg/mL of dimethyl sulphoxide (DMSO) and 0.038  $\mu$ g/mL of trolox, specific scavengers of those radicals respectively. In the deoxyribose assay the second-order rate constant was  $3.56 \times 10^{11} M^{-1} S^{-1}$ , similar to that obtained for some non-steroidal anti-inflammatory drugs. Phycocyanin also inhibits liver microsomal lipid peroxidation ( $IC_{50} = 12$  mg/mL), the CL response of PMNLs ( $p < 0.05$ ) as well as the edema index in GO-induced inflammation in mouse paw ( $p < 0.05$ ).

*Conclusions:* To our knowledge this is the first report of the antioxidant and anti-inflammatory properties of c-phycoerythrin.

**Key words:** Antioxidant – Chemiluminescence – C-Phycocyanin – Blue-green algae – Free radical scavenger

### Introduction

C-phycoerythrin is a protein-bound pigment found in blue-green algae. Phycocyanin monomers are themselves made

up of two distinguishable protein subunits designated  $\alpha$  and  $\beta$ , which contain at least three covalently attached bilin chromophores, open chain tetrapyrroles with no metal complexes [1]. These prosthetic groups account for about 4% of the algae mass, indicating the presence of about sixteen chromophoric groups per unit molecular weight [2]. It occurs in four different structural forms, monomeric, trimeric, hexameric and decameric [3], and is the most abundant pigment in blue-green algae, accounting for more than 20% of algal dry weight [4].

The chemical structure of the bilin chromophores in c-phycoerythrin, (open chain tetrapyrroles) are very close to that of bilirubin. Stocker et al. [5] reported that bilirubin is an antioxidant of possible physiological importance because it could scavenge peroxy radicals by donating a hydrogen atom attached to the C-10 bridge of the tetrapyrrole molecule to form a carbon-centered radical with resonance stabilization extending over the entire bilirubin molecule.

It is well known that reactive oxygen species (ROS) are involved in a diversity of important processes in medicine including, among others: inflammation, atherosclerosis, cancer, reperfusion injury [6]. One way by which a substance can interfere with these processes is by acting as antioxidant or free radical scavenger.

Taking these data into account, we postulated that c-phycoerythrin may be a putative antioxidant and decided to evaluate it in some in vitro and in vivo experimental models.

### Materials and methods

#### Reagents

Luminol and xanthine oxidase (XO 20 U/mL) were obtained from Boehringer Mannheim GmbH (Germany). Trolox and p-iodophenol were from Aldrich Chemical Co. (Milwaukee, WI, USA). Superoxide dismutase (SOD) (3333 U/mL) from bovine erythrocytes and Hypoxanthine (HX) were from Serva Feinbiochimica (Heidelberg, Germany). Tert-butylhydroperoxide (*t*-BOOH) and butyl hydroxytoluene (BHT) were obtained from Sigma Chemical Co. (St. Louis, MO, USA). Sodium dodecylsulphate (SDS), hydrogen peroxide ( $H_2O_2$ ), ascorbic

acid, 2-deoxyribose and thiobarbituric acid (TBA), were from BDH Ltd (Poole, UK).  $\text{FeCl}_3$  and glucose oxidase (GO) (1.4 U/mg) were from Merck (Darmstadt, Germany). All other reagents were of analytical grade.

Phycocyanin was obtained from *Arthrospira maxima* species and purified by the method of Neufeld and Riggs [7].

Phosphate-buffered saline solution (PBS) consisted of NaCl 0.14 M, KCl 2.7 mM,  $\text{Na}_2\text{HPO}_4$  12 mM,  $\text{KH}_2\text{PO}_4$  1.5 mM, CaCl 0.9 mM and  $\text{MgCl}_2$  0.49 mM.

### Animals

Male  $\text{OF}_1$  mice weighing 22–25 g and male Sprague Dawley rats (220–250 g) were purchased from the National Center for Laboratory Animals Production (CENPALAB, Havana, Cuba). The animals were housed under controlled temperature ( $t = 25^\circ\text{C}$ ) and air humidity (60%) with a 12 h light-dark cycle, and kept on a standard laboratory diet and drinking water ad libitum.

### Chemiluminescence measurements

The scavenging action of c-phycocyanin was determined against different types of oxygen radicals, which were generated by specific chemical reactions and detected by LCL.

A well known scavenger for each radical was used as control for the paradigm and to compare its effect with that produced by c-phycocyanin.

Chemiluminescence was measured in millivolts in an LKB Wallac 1250 luminometer coupled to an LKB 2210 two channel recorder.

Superoxide radical scavenging activity was determined as described by Pascual et al. [8]. The reaction consisted of 800  $\mu\text{L}$  of a mixture containing 68 mM glycine buffer pH 8.6, 10  $\mu\text{M}$  luminol and 5  $\mu\text{M}$  p-iodophenol. Fifty  $\mu\text{L}$  of distilled water or c-phycocyanin aqueous solutions were added. Then 2  $\mu\text{L}$  of xanthine oxidase (2 U/mL) were added and the reaction was initiated with 10  $\mu\text{L}$  of hypoxanthine 1 mM. The intensity was registered immediately. SOD was used in the system as a control.

Hydroxyl radicals were generated from the Fenton reaction. The method used was described by Pascual and Romay [9]. In brief, to 800  $\mu\text{L}$  of a mixture containing: 50 mM  $\text{K}_2\text{HPO}_4$ - $\text{KH}_2\text{PO}_4$  buffer pH 7.8, 2 mM EDTA and 0.1 mM luminol, 0.1 mL of distilled water, sample or specific scavenger (DMSO) was added. Then 5  $\mu\text{L}$  of 6 mM  $\text{H}_2\text{O}_2$  was also added and mixed. The reaction was started with 10  $\mu\text{L}$  of 20 mM  $\text{FeSO}_4$  and after rapid mixing, the CL signal was immediately registered.

Determination of alkoxy radical scavenging activity was performed by measuring the inhibition of the CL produced by the reaction of t-BOOH with ferrous ions in the presence of luminol, as previously described [9]. The reaction mixture consisted of: 800  $\mu\text{L}$  of 50 mM glycine buffer pH 8.6, 50 mM SDS, 0.025 mM luminol. Ten  $\mu\text{L}$  of distilled water was added and mixed prior to the addition and mixing of 5  $\mu\text{L}$  of 7.3 mM t-BOOH.

The reaction was started with 10  $\mu\text{L}$  of 0.4 mM  $\text{FeSO}_4$  and immediately after rapid mixing the chemiluminescence signal (mV) was recorded. Trolox, a water soluble analogue of vitamin E, was used as specific scavenger of these radicals.

Double quenching experiments were done in each CL system in order to determine whether the effect of the phycocyanin was due to the scavenging of the desired oxygen free radical or the trapping of other free radical species. These experiments were done by measuring the luminous signal before (Io) and after (I) adding increasing concentrations of c-phycocyanin in the absence and presence (sufficient to cause 50% inhibition) of the specific scavenger (SOD, DMSO or trolox). The slope of the plot is equal to  $k\tau$  where  $\tau$  is the life time of the radical in the absence of phycocyanin. When both phycocyanin and the specific scavenger compete for the same radical, a decrease in the slope must be expected.

### Effect of phycocyanin on the CL response of isolated PMNLs PMNL preparation

Human leukocytes were obtained as described previously [8] from 10 mL heparinized (20 U/mL) venous blood from healthy volunteers, who had not taken any drug during the week before blood sampling. The blood was mixed with an equal volume of an ACD-dextran-glucose mixture consisting of: 1.5 mL acid citrate dextrose (ACD) in 0.9% NaCl (24.5 g glucose/L, 22 g sodium citrate dihydrate/L and 7.3 g citric acid/L), 5 mL of 6% (w/v) Dextran T-500 in 0.9% of NaCl and 3.5 mL of 5% glucose in 0.9% NaCl.

After mixing well and allowing to stand for 45–60 min at room temperature, the upper layer containing the leukocytes was removed by aspiration and three times its volume of ammonium chloride 0.8% added in order to hemolyze the remaining red cells. The cells were centrifuged for 10 min at 500 g at  $4^\circ\text{C}$ . Then, 2 mL of 0.9% NaCl were added, followed by 3 mL of cold distilled water which were mixed and allowed to stand for 2 min, then mixed with 3.6% NaCl and centrifuged for 10 min at 500 g. Finally the cells were resuspended in approximately  $5 \times 10^6$  cells per mL PBS.

### Opsonisation of particles

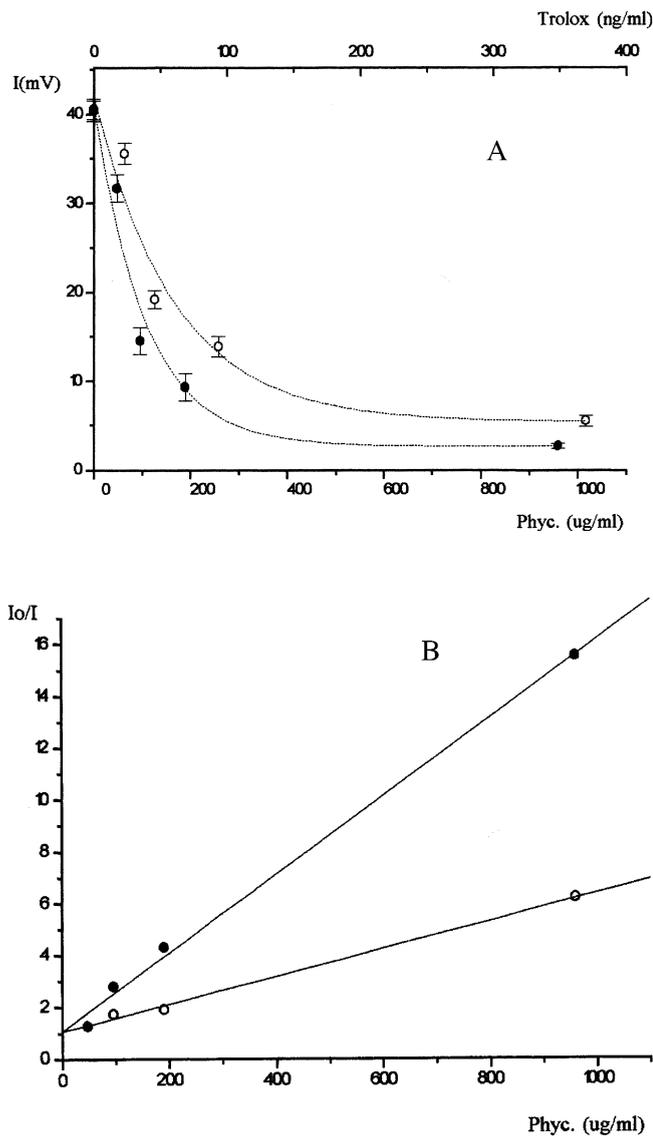
Cells from a fresh culture of *Saccharomyces cerevisiae* were washed and then put into a boiling water bath for 30 min. After washing in saline they were resuspended at a concentration of  $2 \times 10^8$  particles/mL PBS. The opsonisation procedure was carried out by incubating a mixture of 200  $\mu\text{L}$  human serum with 1.8 mL of yeast suspension for 30 min at  $30^\circ\text{C}$  immediately prior to the experiment. A luminol stock solution of  $10^{-2}$  M in DMSO was prepared and was diluted to  $10^{-4}$  M in PBS prior to use.

### Chemiluminescence assay

Chemiluminescence was performed as described previously [8, 10] with minor modifications. Briefly, 200  $\mu\text{L}$  of opsonised yeast cells, 450  $\mu\text{L}$  of phosphate-buffered saline (PBS), 200  $\mu\text{L}$  luminol  $10^{-4}$  M and 50  $\mu\text{L}$  of water or different phycocyanin concentrations, were incubated for 10 min in the measuring cuvette. Just before the assay, 100  $\mu\text{L}$  of an isolated leukocyte suspension were added and the light intensity was measured every 3 min at  $37^\circ\text{C}$ . In a system without cells there was no interaction between luminol and phycocyanin. The viability of PMNLs after being exposed to the higher c-phycocyanin concentration (incubated at  $37^\circ\text{C}$  water bath for 40 min) was assessed by the Trypan blue exclusion test. The viability obtained was 98%.

### Inhibition of damage to 2-deoxyribose

Evaluation of the inhibition of damage to 2-deoxyribose, measured as formation of thiobarbituric acid-reactive material [11], was carried out as an alternative measure of the hydroxyl radical scavenger capacity of c-phycocyanin. Mixtures contained, in a final volume of 1.2 mL: deoxyribose 2.8 mM,  $\text{KH}_2\text{PO}_4$ /KOH buffer 15 mM pH 7.4,  $\text{FeCl}_3$  20  $\mu\text{M}$ , EDTA 100  $\mu\text{M}$ ,  $\text{H}_2\text{O}_2$  2.8 mM and ascorbic acid 100  $\mu\text{M}$ .  $\text{FeCl}_3$  and ascorbate solutions were made up in bidistilled water just before use.  $\text{FeCl}_3$  and EDTA were premixed prior to addition to the reaction mixture. Ascorbic acid was added in order to start the reaction. Reaction mixtures were incubated at  $37^\circ\text{C}$  for 1 h. After addition of 1 mL of TBA 1% (w/v) in 0.05 mM NaOH and 1 mL of TCA 2.8% in water, the mixture was heated at  $100^\circ\text{C}$  for 20 min. The pink chromogen that progressively developed was then measured at 532 nm after cooling, against appropriate blanks. The second-order rate constants  $k_s$  were calculated using the data obtained in the presence of EDTA from the slope of a plot of  $1/A_{532\text{nm}}$  against the test compound concentration.

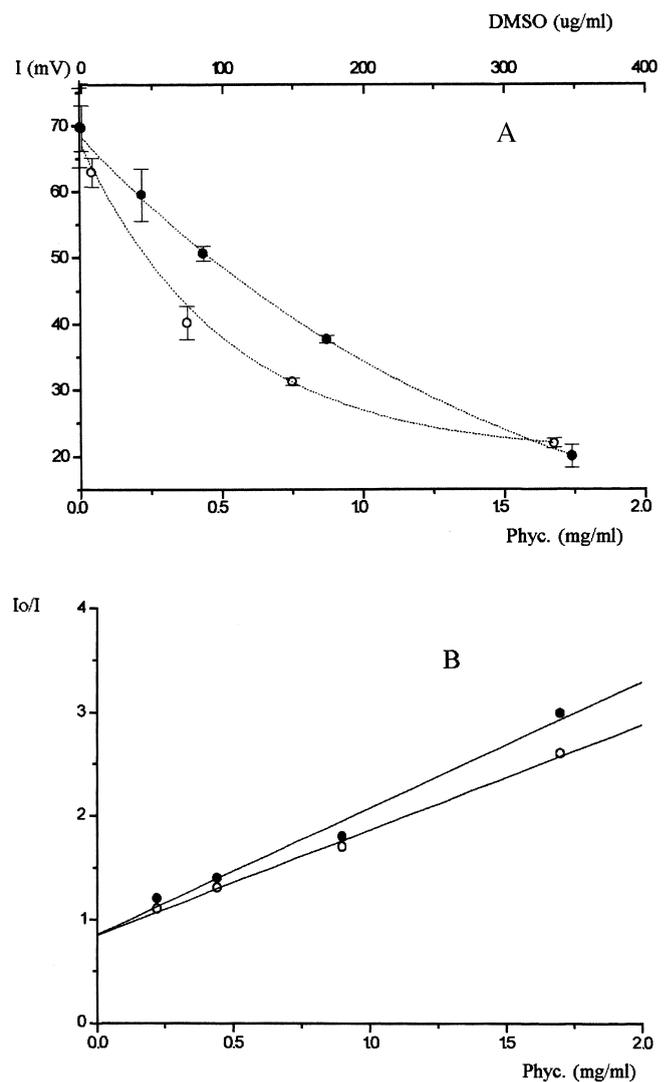


**Fig. 1.** Effect of c-phycoyanin on chemiluminescence intensity produced by alkoxy radicals generated by the reaction of tert-butylhydroperoxide with ferrous ion. (A) Decrease in Chemiluminescence signal (mV) with increasing c-phycoyanin (●) or trolox (○) concentration. Each point represents the mean of three determinations. Vertical bars represent  $\pm 1$  SD. (B) Double quenching experiment carried out to determine the chemiluminescence signal before ( $I_0$ ) and after ( $I$ ) adding increasing concentrations of c-phycoyanin in the absence (●) and presence (○) of trolox (46  $\text{ng/ml}$ ). Each point represents the mean  $I_0/I$  value obtained from three determinations.

#### *Inhibition of liver microsomal lipid peroxidation induced by $\text{Fe}^{+2}$ -ascorbic acid*

Rat liver microsomes were prepared by differential ultracentrifugation as previously described [12] and stored at  $-80^\circ\text{C}$  until use. Protein was assayed by the Lowry method [13].

The procedure was carried out as described [14]. The microsomes (final concentration 1.3  $\text{mg}$  protein/ml) were incubated at  $37^\circ\text{C}$  in Tris buffer 50  $\text{mM}$  pH 7.4 before induction of lipid peroxidation with 10  $\mu\text{M}$   $\text{FeSO}_4$  and freshly prepared 0.2  $\text{mM}$  ascorbic acid. The reaction was stopped by adding 0.3 mL of the incubation mixture to 2 mL of ice-cold TBA-TCA-HCl-BHT. After heating for 15 min at  $80^\circ\text{C}$  and centrifugation



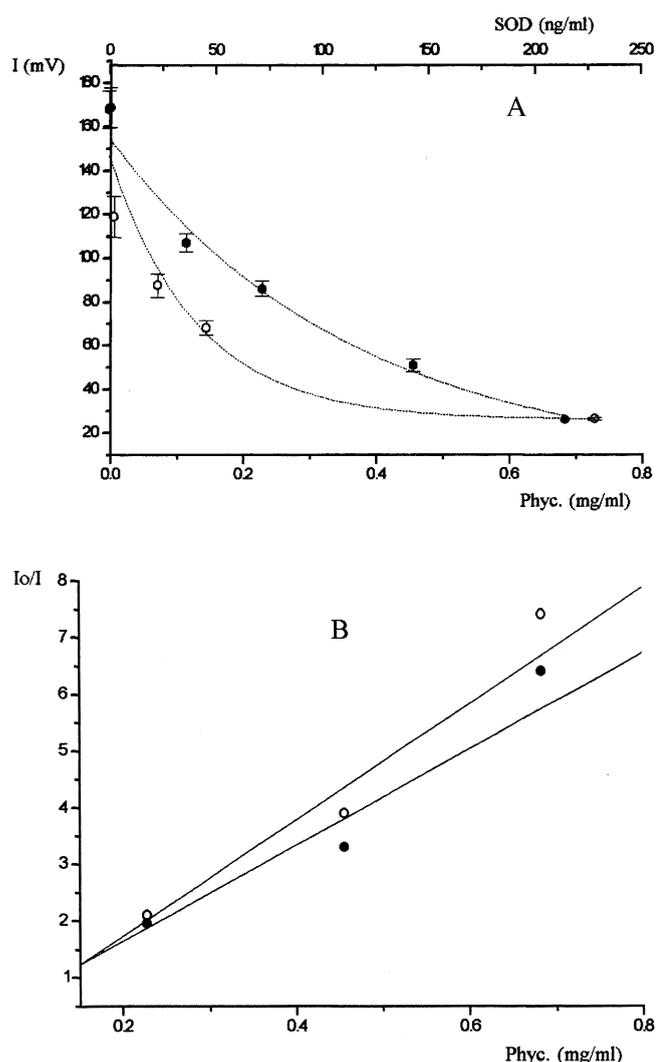
**Fig. 2.** Effect of c-phycoyanin as scavenger of hydroxyl radicals produced by the Fenton reaction. (A) Decrease in chemiluminescence signal (mV) with increasing c-phycoyanin (●) or DMSO (○) concentrations. Each point represents the mean of three determinations. Vertical bars represent  $\pm 1$  SD. (B)  $I_0/I$  vs c-phycoyanin concentration plot obtained as in Figure 2 in the absence (●) and presence (○) of DMSO (149  $\mu\text{g/ml}$ ). Each point represents the mean  $I_0/I$  value obtained from three determinations.

for 15 min at 2000 g, the absorbance at 535 nm was determined. The TBA-TCA-HCl solution was prepared by dissolving 41.6  $\text{mg}$  TBA/10 mL TCA (16.8% w/v in 0.125 N HCl). To 10 mL TBA-TCA-HCl, 1 mL of BHT (1.5  $\text{mg/ml}$  ethanol) was added.

All the spectrophotometric measurements were done in a Spekoll 220 from Carl Zeiss (Jena, Germany).

#### *GO-induced inflammation in mouse paw*

The animal model used was described by Spillert et al. [15]. Phycocyanin (50, 100 or 200  $\text{mg/Kg}$  in saline) or DMSO (1  $\text{g/Kg}$ ), as positive control, were administered orally and i.p., respectively, to male OF1 mice. One hour later, the mice were injected in the right hind foot with 50  $\mu\text{L}$  of physiological saline and in the left foot with 50  $\mu\text{L}$  of 100 U/mL GO. The animals were killed at 1.5 h post injection, both hind feet amputated at the tibiotarsal joint and each paw weighed. The



**Fig. 3.** Effect of various c-phycoyanin concentrations on chemiluminescence produced in the luminol-XO-HX system. (A) Decrease in chemiluminescence signal (mV) with increasing c-phycoyanin (●) or SOD (○) concentrations. Each point represents the mean of three determinations. Vertical bars represent  $\pm 1$  SD. (B)  $I_o/I$  vs c-phycoyanin concentration plot obtained as in Figure 2 in the absence (●) and presence (○) of SOD (23 ng/mL). Each point represents the mean  $I_o/I$  value from three determinations.

difference in weight of hind paws of each animal was called the edema index (EI).

#### Statistical analysis

One-way ANOVA followed by Duncan's multiple comparison test were used to calculate the significance of the differences between the means. The  $IC_{50}$  was calculated using a GraphPad InPlot software (GraphPad Software Inc., version 4.03, 1992).

## Results

### Chemiluminescent measurements

The CL produced by the reaction of t-BOOH with ferrous

**Table 1.** Effect of phycocyanin on the chemiluminescence response of human leukocytes.

	AUC $\bar{X} \pm SD$	Inhibition %
Control	6662 $\pm$ 221.3 <sup>a</sup>	–
Phycocyanin 1 mg/mL	5800 $\pm$ 246.5 <sup>b</sup>	12.9
2 mg/mL	4647 $\pm$ 227.0 <sup>c</sup>	30.2
3 mg/mL	3841 $\pm$ 296.5 <sup>d</sup>	42.3

Different letters:  $p < 0.05$  vs. each other,  $n = 5$ .

**Table 2.** Effect of phycocyanin on GO-induced inflammation in the mouse paw.

Treatment	Edema index (g)	Inhibition %
DMSO (1 g/Kg)	0.045 $\pm$ 0.007 <sup>a</sup>	64.05
Phycocyanin 50 mg/Kg	0.110 $\pm$ 0.010 <sup>b</sup>	4.43
100 mg/Kg	0.080 $\pm$ 0.007 <sup>c</sup>	35.46
200 mg/Kg	0.061 $\pm$ 0.009 <sup>d</sup>	51.05
Glucose oxidase 100 U/mL	0.125 $\pm$ 0.008 <sup>b</sup>	–

Different letters:  $p < 0.05$  vs. each other,  $n = 7$  mice per group.

Phycocyanin was administered orally 1 h before GO. The oedema was measured 1.5 h after injection of GO into the paw. The oedema index indicates the difference in weight between the hind paws.

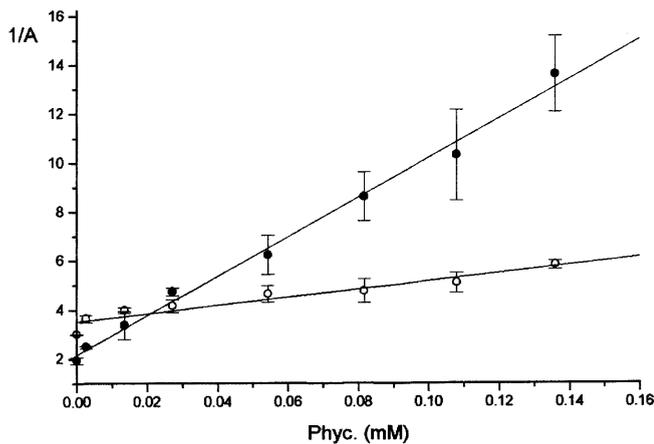
ion was used for evaluation of alkoxy radical scavenging capacity of c-phycoyanin. The results show that c-phycoyanin inhibited the CL in this system (Fig. 1A). A comparison with trolox, a water soluble analogue of vitamin E, indicates that 0.038  $\mu$ g/mL of trolox causes approximately the same effect as 76  $\mu$ g/mL of c-phycoyanin in terms of 50% inhibition of the CL produced in this system.

C-phycoyanin was evaluated as a scavenger of hydroxyl radicals by determining the inhibition of CL produced by the Fenton reaction with luminol. As shown in Figure 2A, the chemiluminescence signal (mV) was inhibited by increasing phycocyanin concentrations. In this system, 0.91 mg/mL of phycocyanin caused the same inhibition (50%) as 0.125 mg/mL of DMSO, which was used as control.

In the HX-XO chemiluminescence system (Fig. 3A), phycocyanin inhibited the signal in a dose-dependent manner. However, when the HX-XO reaction was used to reduce nitroblue tetrazolium (NBT) dye, no inhibitory effect was observed (data not shown). In the double quenching experiments (Fig. 1B and 2B), the slope decreased when both phycocyanin and the specific scavenger were added together, indicating competition for the radical generated. This behaviour was not observed in the  $O_2^-$ -generator system (Fig. 3B) in which the slope is independent of the previous addition of the specific scavenger (SOD) and is even increased.

### Effect of phycocyanin on the CL response of PMNLs

Human leukocytes stimulated with opsonized yeast cells produced a typical time-dependent CL response with a maximal intensity at approximately 16 min followed by a decline. Table 1 shows the means of the areas under the curve (AUC) obtained for each phycocyanin concentration



**Fig. 4.** Plots of  $1/A_{532\text{nm}}$  against concentration of c-phycoerythrin using data obtained from deoxyribose oxidation in the presence (●) or absence (○) of EDTA. Values for each point are means of three experiments. Vertical bars represent  $\pm 1$  SD.

and control. The CL signal was significantly reduced with respect to control by increasing phycoerythrin concentrations. Significant statistical differences were also obtained among phycoerythrin concentrations.

#### Deoxyribose assay

In the deoxyribose assay, a more conventional method, phycoerythrin inhibited the deoxyribose damage in a concentration-dependent fashion. The  $IC_{50}$  calculated for c-phycoerythrin was 0.86 mg/mL. In this system, a typical competition profile was obtained (Fig. 4). This was suitable for the calculation of a second-order rate constant of  $3.56 \times 10^{11} \text{ M}^{-1} \text{ S}^{-1}$ , considering a molecular weight of 36 700 g/mol for c-phycoerythrin monomer [16].

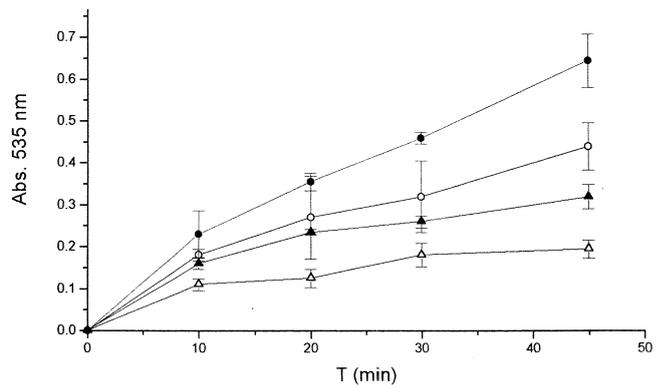
#### Microsomal lipid peroxidation inhibition

Addition of c-phycoerythrin (8, 12, 20 mg/mL) to isolated microsomes in the presence of  $\text{Fe}^{+2}$ -ascorbate, resulted in a concentration dependent decrease in lipid peroxidation ( $IC_{50} = 12 \text{ mg/ml}$ ). Both the rate and the final extent of lipid peroxidation were reduced by adding c-phycoerythrin (Fig. 5).

In the presence of iron alone, c-phycoerythrin did not cause lipid peroxidation, indicating that it does not have pro-oxidant effect in this system. Furthermore, its inhibitory effect was exerted during the incubation time, since it had no effect when it was added with the TBA reagent.

#### GO-induced inflammation in mouse paw

Differences in weight between the hind paws of animals in the various groups are shown in Table 2. Phycoerythrin at doses of 100 and 200 mg/Kg orally was able to inhibit significantly peroxide-induced inflammation in a dose dependent fashion with a  $ED_{50}$  of  $170.3 \pm 1.62 \text{ mg/Kg}$ . DMSO was used as a positive control and it also inhibited the inflammatory response induced by GO in mouse paw.



**Fig. 5.** Time course of microsomal non-enzymatic lipid peroxidation as affected by various concentrations of c-phycoerythrin: (●) no addition, (○) 8 mg/mL, (▲) 12 mg/mL, (△) 20 mg/mL. Values for each point are means of three determinations. Vertical bars represent  $\pm 1$  SD.

#### Discussion

In this study, we have applied established *in vitro* and *in vivo* assays in order to evaluate the antioxidant action of c-phycoerythrin. This natural product was able to scavenge alkoxy and hydroxyl radicals. Two methods were used to evaluate hydroxyl radical scavenging by c-phycoerythrin because hydroxyl radical is one of the most potent oxidizing species and its extreme reactivity naturally poses problems with regard to its detection [17]. In both methods, inhibition was observed at relatively high concentrations of the product.

Phycoerythrin also quenched the CL signal generated in the HX-XO system, but this effect can not be ascribed to  $\text{O}_2^-$  scavenging, as demonstrated by double-quenching and NBT reduction assays. One possible explanation for this behaviour is that phycoerythrin quenches CL by binding to an intermediate or co-oxidising species that may be involved in the CL reaction [18].

The inhibitory effect observed on microsomal lipid peroxidation most probably is due to a metal-binding capacity of c-phycoerythrin, since chain-breaking antioxidants often introduce a lag period into the peroxidation process, corresponding to the time taken for the antioxidant to be consumed, whereas metal-binding antioxidants will give a constant inhibition throughout the reaction [19]. Another indication of such an action, is the ability of c-phycoerythrin to inhibit deoxyribose damage in a site-specific manner (in the absence of EDTA) [19]. Further experiments must be performed to obtain more evidence for the ability of phycoerythrin to chelate metal ions.

In the deoxyribose assay, a second-order rate constant calculated for phycoerythrin was similar to that obtained, by the same method, for some non-steroidal anti-inflammatory drugs, such as indomethacin and ibuprofen ( $1.8 \times 10^{10} \text{ M}^{-1} \text{ S}^{-1}$ ) [20].

Chemiluminescence of PMNLs is the final result of luminol oxidation by strong oxidants, such as oxygen radicals and peroxides, emanating from enzymatic reactions. For addition to the myeloperoxidase- $\text{H}_2\text{O}_2$ -halide system, the release of arachidonic acid by phospholipase  $A_2$  and of diacylglycerol and inositol trisphosphate by phospholipase C, the metabolism of arachidonic acid by the

cyclooxygenase and lipoxygenase pathways, the activation of membrane NADPH oxidase by diacylglycerol and calcium mobilization by inositol trisphosphate are all able to induce the CL reaction. Inhibition of any of these mechanisms suppresses the CL response [21].

Phycocyanin was able to inhibit the LCL in a dose-dependent fashion, most probably through its capacity to scavenge free radicals ( $\text{OH}^\bullet$ ,  $\text{H}_2\text{O}_2$ ,  $\text{RO}^\bullet$ ) and peroxides arising during the respiratory burst of phagocytic cells. However, it is also possible that phycocyanin could diminish CL signals in other ways, e.g. by affecting enzymes involved in the production of reactive oxygen species by activated phagocytes, NADPH oxidase and myeloperoxidase, or by interfering either with the binding of the stimulant or the arachidonic acid metabolism pathway. In this regard, we have recent evidence for inhibition by phycocyanin of  $\text{LTB}_4$  release in an animal model of inflammation (manuscript in preparation).

The peroxide-induced inflammatory response is a valuable *in vivo* model in order to test agents with potential scavenging effect against  $\text{H}_2\text{O}_2$  and  $\text{OH}^\bullet$ . GO injected into the mouse paw reacts with endogenous glucose and generates  $\text{H}_2\text{O}_2$  which subsequently produces  $\text{OH}^\bullet$  radicals; both together are responsible for tissue damage and for the accompanying inflammatory changes [15]. Phycocyanin reduced the edema produced by glucose oxidase in the mouse paw. This anti-inflammatory effect must be due, at least in part, to the scavenging of hydroxyl radicals, taking into account the fact that DMSO, a well known scavenger of  $\text{OH}^\bullet$  radicals, also inhibited the inflammatory response induced by GO.

Currently, there is a consensus that much of the damage induced by  $\text{H}_2\text{O}_2$  *in vivo* is due to its conversion to highly-reactive oxidants, mainly  $\text{OH}^\bullet$  [19]. Therefore, the scavenging action of phycocyanin against  $\text{OH}^\bullet$  is probably relevant to its anti-inflammatory effects.

Very recently, research carried out in our laboratory has confirmed the anti-inflammatory effects of phycocyanin in other experimental models of inflammation such as cotton pellet granuloma in the rat and TPA-induced inflammatory response in the mouse ear (manuscript in preparation).

Finally, to our knowledge, this is the first time that both antioxidant and anti-inflammatory properties have been described for c-phycocyanin.

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